

This is a refereed journal and all articles are professionally screened and reviewed

ORIGINAL ARTICLE

The Screening of Phytoconstituents, Antibacterial and Antifungal Activities of *Brysocarpus Coccineus* Schum and Thonn. Stem (Connaraceae).

¹Abdulmumeen A. HAMID and ²Olapeju O. AIYELAAGBE

¹Department of Chemistry, University of Ilorin, Nigeria.

Abdulmumeen A. HAMID and Olapeju O. AIYELAAGBE: The Screening of Phytoconstituents, Antibacterial and Antifungal Activities of *Brysocarpus Coccineus* Schum and Thonn. Stem (Connaraceae).

ABSTRACT

The uses of *Brysocarpus coccineus* stem in traditional African medicine have led to the antibacterial and antifungal activities of the hexane, ethylacetate and methanol stem extracts of the plant evaluation using six pathogenic bacteria and six fungi. The extracts inhibited the 12 test organisms to different degrees. Hexane, ethylacetate and methanol successive extracts of *Brysocarpus coccineus* stem effectively inhibited the growth of *Escherichia coli*, *Bacillus subtilis* and *Pseudomonas aeruginosa* at concentrations between 12.5 and 200mg/ml, while the extracts showed lower inhibition on *Staphylococcus aureus*, *Salmonellae typhii and Klebsiellae pneumonae*. The sensitivity of test bacteria was concentration dependent, activity being higher at higher concentrations of the three extracts. Ethylacetate and methanol exhibited higher antifungal properties on *Rhizopus stolon* and *Epidermophyton floccosum*, *Tricophyton rubrum* and *Aspergillus niger* with activity comparable to that of the reference drug tioconazole. Preliminary phytochemical screening of the three extracts revealed the presence of saponins, reducing sugar, steroids, glycosides, flavonoids and anthraquinones.

Key words:

Introduction

Medicinal plants have played essential roles since the early stage of life in the treatment of all kinds of diseases. African flora is very rich in these medicinal plants. The bacterial and fungal infections are also prevalent in our society and this leads to various diseases like gonorrhea, syphilis, pneumonia and different skin disorders. The search for new, safer and cheaper drugs especially from plants, to treat and cure these diseases is on the increase. *Brysocarpus coccineus* Schum and Thonn. (Connaraceae) is a climbing shrub found in Africa [1]. The plant especially its leaf is used in traditional medicine for the treatment of veneral diseases, impotency, diarrhoea, jaundice, piles, dysentery, earache, sore

of mouth and skin, tumour, wounds, stomatitis, rheumatism, swellings and urinary disorders [2,3,4,8]. The pharmacological properties of *Brysocarpus coccineus* as antioxidant, anti-inflammatory, analgesic, antidiarrhoeal and antipyretic have been established [5,6,7,8,9].

Further, the uterotonic, molluscidal, hepatoprotective, anxiolytic and sedative activities of various extracts of the plant have also been reported [10,11,12,13]. A coumaroyl derivative, 4-hydroxycoumarin, flavonoids and flavones have been isolated from the leaves of *Brysocarpus coccineus* by Vickery and Vickery [14]. The presence of quercetin, quercetin 3-O- α -arabinose and quercetin 3-O- β -D-glucose from the bioactive ethylacetate and n-butanol soluble parts of ethanol extracts of *Brysocarpus coccineus* was also

²Department of Chemistry, University of Ibadan, Nigeria.

established by Ahmadu et al.,[15].

From the previous work, no scientific investigations have been reported on biological activities of the stem of *Brysocarpus coccineus*. Hence we now study the preliminary phytoconstituents, antibacterial and antifungal activities of *Brysocarpus coccineus* stem.

Materials and methods

Collection and Authentication of the Plant Material:

The plant material of *Brysocarpus coccineus* was collected from Ibadan area of Oyo State, Nigeria, in the month of November 2009. Botanical identification and authentication was done by Mr. A.W. Ekundayo of the Forestry Research Institute of Nigeria (FRIN), Ibadan where a voucher specimen was deposited with the herbarium file number FHI108801.

Preparation of Plant Extracts:

The whole plant of *Brysocarpus coccineus* was separated into leaves and stem, air-dried and weighed (stem, 1100g and leaves, 534g). The dried stem was successively extracted in hexane, ethylacetate and methanol for 10 days respectively using cold extraction method. The resultant hexane (6g), ethylacetate (9g) and methanol (8g) extracts were obtained by evaporation and stored in the refrigerator for further use.

Phytochemical Studies:

The Preliminary phytochemical screening of the hexane, ethylacetate and methanol extracts of *B. coccineus* stem was done using standard procedures [16,17,18,19,20].

Antimicrobial Assay:

Microorganisms:

Cultures of six human pathogenic bacteria made up of four gram negative and two gram positive were used for the antibacterial assay. Namely; Salmonella typhii (UCH 4801), Escherichia coli (UCH 00260), Pseudomonas aeruginosa (UCH 1102) and Klebsiellae pneumonae(UCH 2894) (gram negative), Bacillus subtilis (UCH 74230) and Staphylococcus aureus (UCH 2473) (gram positive). For the Antifungal assay, six fungi were also utilized. These were; Candida albicans, Aspergillus niger, Rhizopus stolon, Penicillum notatum, Tricophyton rubrum and Epidermophyton floccosum. All the

microorganisms used were clinical strains from the Department of Medical Microbiology (University College Hospital, Ibadan) and screened in the Laboratory of Pharmaceutical Microbiology Department, University of Ibadan, Ibadan, Nigeria.

Media:

Nutrient agar, Sabouraud dextrose agar, nutrient broth and tryptone soya agar were used in this study. Hexane, ethylacetate and methanol were also used in solubilizing the extracts and as negative controls in the assays.

Antimicrobial Agents:

Gentamycin $(10\mu g/ml)$ and Tioconazole (0.7mg/ml) were included as standard reference drugs in the study.

Antimicrobial Activity Determination:

Agar Diffusion-pour Plate Method (Bacteria):

An overnight culture of each organism was prepared appropriately from its stock and inoculated each into the sterile nutrient broth of 5ml, each incubated for 18-24hrs at 37°C. From overnight culture, 0.1ml of each organism was taken and put into the 9.9mls of sterile distilled water to get (1:100) 10⁻² of the dilution of the organism.

From the diluted organism (10°2), 0.2ml was taken into the prepared sterile nutrient agar cooled to about 40-45°C, then poured into sterile Petri dishes and allowed to solidify for about 45-60mins. Using a sterile cork-borer of 8mm diameter, the wells were made according to the number of the test tubes for the experiment. For this work 8 wells were made. The graded concentrations of the extracts were put into the wells accordingly including the controls. The studies were done in duplicates to ascertain the results obtained. The plates were left on the bench for about 2hrs to allow the extract diffuse properly into the nutrient agar i.e. pre-diffusion. The plates were incubated for 18-24hrs at 37°C.

Agar Diffusion-surface Plate Method (Fungi):

A sterile Sabouraud dextrose agar was prepared accordingly and aseptically poured into the sterile plates in duplicates and solidified properly. 0.2ml of the (1:100) 10⁻² of the organism was spread on the surface of the agar using a sterile Petri-dish cover to cover all the surface of the agar. Eight wells were bored using a sterile cork-borer of 8mm diameter. The graded

concentrations of the extracts were put into the well accordingly including the controls. All the plates were left on the bench for 2hrs to allow the extract diffuse properly into the agar i.e. prediffusion. The plates were incubated at 25°C for 72hrs [21,22,23].

Results and discussions

The results of the phytochemical screening of the hexane, ethylacetate and methanol extracts of *Brysocarpus coccineus* stem are presented in Table1. Preliminary phytochemical screening revealed the presence of reducing sugar, steroids, glycosides, flavonoids and anthraquinones in all the extracts. There was presence of saponins in ethylacetate and methanol extracts of *Brysocarpus coccineus* stem but absent in hexane extract of the plant. However, tannins and alkaloids were not found in all the three extracts of *Brysocarpus coccineus* stem.

The result of antibacterial activities of the hexane, ethylacetate and methanol extracts at concentrations ranging from 6.25 to 200mg/ml was presented in Table 2.

The bacteria used were clinical strains of Staphylococcus aureus and Bacillus subtilis (gram positive), Escherichia coli, Pseudomonas aeruginosa, klebsiellae pneumonae and salmonellae typhii (gram negative).

All the bacteria strains were sensitive to the three extracts at concentrations between 12.5 and 200mg/ml, but hexane extract exhibited higher inhibition on *Escherichia coli*, *Bacillus subtilis* and *Pseudomonas aeruginosa* than both ethylacetate and methanol extracts of *Brysocarpus coccineus*. Meanwhile, all extracts also inhibited

Absent

present

the growth of *Staphylococcus aureus* and *salmonellae typhii*, while the inhibition of *klebsiellae pneumonae* by the extracts was very low.

Further, the sensitivity of the test bacteria to all the extracts were concentration dependent, activity being higher at higher concentrations of the extracts.

The result of the antifungal activities of the hexane, ethylacetate and methanol extracts at concentrations ranging from 6.25 to 200mg/ml was presented in Table 3. six clinical strains of fungi were used in our study. candida, albicans, Aspergillus niger, Rhizopus stolon, penicillum notatum, Tricophylonrubrum and epidermophyton floccosum. The six test fungi were sensitive to all extracts.

Hexane, ethylacetate and methanol extracts of the stem of *Brysocarpus coccineus* exhibited higher antifungal properties on *Aspergillus niger, Rhizopus stolon, penicillum notatum, Tricophyton rubrum* and *Epidermophyton floccosum* with activity comparable to that of the reference drug tioconazole against *Rhizopus stolon, Tricophyton rubrum, Epidermophyton floccosum and Aspergilus niger* for hexane extract, and *Rhizopus stolon* and *Epidermophyton floccosum* for both ethylacetate and methanol extracts.

However, all extracts showed lower inhibition on *Candida albicans*.

Table 1: Phytochemical constituents of the hexane, ethylacetate and methanol extracts of Brysocarpus coccineus stem.

Secondary metabolites	Extracts (whole plant)							
	Hexane	Ethylacetate	Methanol					
Alkaloids	-	-	-					
Saponins	-	++	++					
Tannins	-	-	-					
Reducing sugars	++	++	++					
Steroids	++	++	++					
Glycosides	++	++	++					
Flavonoids	++	++	++					
Anthraquinones	++	++	++					

Table 2: Antibacterial activities of the hexane, ethylacetate and methanol extracts of Brysocarpus coccineus stem.

Extracts	Extract conc/Ref./ Control (mg/ml)		f well = 8 mm f zone of inhibiti	n ition of bacteria(mm)				
		S.a	E.coli	B.sub	Ps.a	Kleb	Sal.	
Hexane	6.25	-	-	-	-	-	-	
	12.5	-	10	-	-	-	-	
	25	-	12	12	10	-	-	

Table 2: Continue.

Table 2. Cont	muc.							
	50	10	14	14	12	-	12	
	100	12	16	16	14	-	14	
	200	14	24	20	18	10	16	
	Hexane	-	-	-	-	-	_	
	Gentamycin	36	34	34	36	34	36	
Ethylacetate	6.25	-	-	-	-	-	-	
•	12.5	-	-	-	-	-	_	
	25	-	10	12	10	-	-	
	50	-	12	14	12	-	_	
	100	10	16	16	14	-	12	
	200	14	18	18	16	10	14	
	Ethylacetate	-	-	-	-	-	-	
	Gentamycin	36	34	34	36	34	36	
Methanol	6.25	-	-	-	-	-	-	
	12.5	-	10	-	-	-	-	
	25	-	12	12	10	-	-	
	50	10	14	14	12	-	10	
	100	12	16	16	14	-	12	
	200	14	20	18	16	10	14	
	Methanol	-	-	-	-	-	-	
	Gentamycin	36	34	36	36	34	34	

Table 3:	Anfungal	activities	of	the	hexane,	ethylacetate	and	methanol	extracts	of	Brysocarpus	coccineus	stem.
----------	----------	------------	----	-----	---------	--------------	-----	----------	----------	----	--------------------	-----------	-------

Extracts	Extract conc/Ref./ Control (mg/ml)	Diameter of well = 8 mm Diameter of zone of inhibition of bacteria(mm)							
		C.a	A.n	Rhiz	Pen	T.r	E.f		
Hexane	6.25	-	-	_	-	-	-		
	12.5	-	-	10	-	-	-		
	25	-	-	12	10	-	-		
	50	10	10	16	12	10	10		
	100	12	12	18	14	12	16		
	200	14	18	20	16	18	20		
	Hexane	-	-	_	_	-	-		
	Tioconazole	28	22	26	24	22	22		
Ethylacetate	6.25	-	-	_	-	-	-		
	12.5	-	-	-	_	-	-		
	25	-	-	10	_	-	10		
	50	-	10	14	10	10	12		
	100	10	12	18	12	12	16		
	200	12	15	24	16	14	20		
	Ethylacetate	-	-	-	_	-	-		
	Tioconazole	24	24	23	24	23	22		
Methanol	6.25	-	-	-	-	-	-		
	12.5	-	-	10	_	-	-		
	25	-	-	12	10	-	-		
	50	10	10	14	12	10	10		
	100	12	12	18	14	12	14		
	200	14	16	26	16	16	18		
	Methanol	-	-	-	-	-	-		
	Tioconazole	24	24	24	24	24	22		
	Key								

	ikey
S.a	Staphylococcus aureus
E.coli	Escherichia coli
B.sub	Bacillus subtilis
Ps.a	Pseudomonas aeruginosa
Kleb	Klebsiellae pneumonae
Sal	Salmonellae typhii
C.a	Candidas albicans
A.n	Aspergillus niger
Rhiz	Rhizopus stolon
Pen	Penicillum notatum
T.r	Tricophyton rubrum
E f	Enidermonhyton floccosum

Conclusions

The antibacterial and antifungal activities of all extracts (hexane, ethylacetate and methanol)

further confirm the ethnomedicinal use of *Brysocarpus coccineus* to treat veneral diseases, diarrhea, rheumatism, impotency, mouth and skin sores, urinary disorder etc. [2,3,4,8].

Acknowledgements

The authors appreciate Mr. Festus of the Department of Pharmaceutical Microbiology, University of Ibadan and Mr. Wale Ekundayo of Forestry Research Institute of Nigeria (FRIN) Ibadan for the assistance provided during this research work.

References

- 1. Dalziel, J.M., 1955. The useful plants of West Tropical Africa. A crown agent for oversea publication London, pp: 568-570.
- Burkill, H.M., 1985. Useful plants of west tropical Africa Ed.2, 1, families A-D, Royal Botanic Gardens, kew.
- Adjanohoun, E.J., A.M.K. Ahyi,, Ake L. Assi, K. Akpagana, P. Chibon, A. El-Hadji, J. Eyme, M. Garba, J.N. Gassita, M. Gbeassor, E. Goudote, S. Guinko, K.K. Houdote, P. Houngnon, A. Keita, A. Keoula, W.P. Klugoocloo, I. Lo, K.M. Siameri, K.K. Tafferne, 1986. Contribution auxetudes ethnobotaniques et floristiques an togo. Agence de cooperation culturelle et Techinique. Paris.
- 4. Irvine, F.R., 1961. Woody plants of Ghana. An oxford university press publication, London, pp. 568-570.
- Akindele, A.J., O.O. Adeyemi, 2007^a. Antiinflammatory activity of the aqueous leaf extracts of Brysocarpus coccineus, *Fitoterapia*, 78: 25-28.
- 6. Oke, J.M., M.O.Hamburger, 2002.screening of some Nigerian medicinal plant for antioxidant activity using 2-2-Diphenyl-picryl-hydrazyl radical. *African journal of biomedical research*, 5(1-2): 77-79
- 7. Akindele, A.J., O.O. Adeyemi, 2006^b. Evaluation of the antidiarrheal activity of brysocarpus coccineus. *Journal of ethnopharmacology*, 108: 20-25.
- 8. Akindele, A.J., O.O. Adeyemi, 2006^a. Analgesic activity of the aqueous leaf extract of bryosocarpus coccineus. *Nigerian journal of health and biomedical sciences*, 5(1): 43-46.
- 9. Amos, S., L. Binda, O.F. Kunle, C. Wambebe, K. Gamaniel, 2002. Uterotonic activity of the ethanolic extract of *Brysocarpus coccineus*. *Pharmaceutical Biology*, 40: 33-38

- 10. Akindele, A.J., O.O. Adeyemi, 2007^b. Antipyretic activity of *Brysocarpus coccineus* schum and thonn(connaraceae). *International Journal of pharmacology*, 3(4): 357-361.
- 11. Okunji, C.O., M.M. Iwu, 1988. Control of Schistosomiasis using Nigerian Medicinal plants as molluscicides. *Int. J. Crude Drug Res.*, 26(4): 246-252.
- 12. Akindele, A.J., K.O. Ezenwanebe, C.C. Anunobi, O.O. Adeyemi, 2010. Hepatoprotective and invivo antioxidant effects of Brysocarpus coccineus Schum and Thonn. (Connaraceae). *Journal of ethnopharmacology*, 129: 46-52.
- 13. Akindele, A.J., O.O. Adeyemi, 2010. Anxiolytic and sedative effects of *Brysocarpus coccineus* Schum and Thonn. (Connaraceae) extract. *International journal of applied research in natural plants.*, 3(1): 28-36
- 14. Vickery, M., B.Vickery, 1980. Coumarins and related compounds in members of the connaraceae). *Toxicological letter*, 5: 115-118
- Ahamadu, A.A., H.S. Hassan, M.U. Abubakar, I.N. Akpulu, 2007. Flavonoids glucosides from *Brysocarpus coccineus* leaves Schum and Thonn. (Connaraceae). *Africa journal of* traditional, complimentary and alternative medicines., 4(3): 257-260.
- 16. Trease, G.E., W.C. Evans, 1983. Drugs of Biological origin. In: pharmacognosy 12th ed. United kingdom: Balliere tindall., pp: 305-540.
- 17. Trease, G.E., W.C. Evans, 1989. pharmacognosy. Brailliar Tiridel can, 13th ed. Macmillan publishers.
- 18. Harborne, J.P., 1991. Phytochemical methods: 2nd ed.p.51, Oxford university press, London.
- Harborne, J.P., 1998. Phytochemical methods: A guide to modern technique of plants analysis,2nd ed. London: Chapman and Hall.
- Edeoga, H.O., D.E. Okwu, B.O. Mbaebie, 2005. Phytochemical constituents of some Nigerian medicinal plants. *African journal of biotechnology*, 4: 685-688.
- 21. Kavanagh, F., 1972. Analytical microbiolology. Academic press, New York.
- 22. Ziwadyk, P., 1972. Enteriobacteriaceae in Zinsser microbiology, 20th ed. pp: 544-546. Georg Thiene verlag, Stuttgart.
- Van den Berge A.D., A.J. Vlietinck, 1991. Methods in plants biochemistry. Vol.6. Assays for bioactivity. Hostettmann K. Ed. Academic press, London, 47-69.